ATP-dependent and ATP-independent Roles for the Rad54 Chromatin Remodeling Enzyme during Recombinational Repair of a DNA Double Strand Break*

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The efficient and accurate repair of DNA double strand breaks (DSBs) is critical to cell survival, and defects in this process can lead to genome instability and cancers. In eukaryotes, the Rad52 group of proteins dictates the repair of DSBs by the error-free process of homologous recombination (HR). A critical step in eukaryotic HR is the formation of the initial Rad51-single-stranded DNA presynaptic nucleoprotein filament. This presynaptic filament participates in a homology search process that leads to the formation of a DNA joint molecule and recombinational repair of the DSB. Recently, we showed that the Rad54 protein functions as a mediator of Rad51 binding to single-stranded DNA, and here, we find that this activity does not require ATP hydrolysis. We also identify a novel Rad54-dependent chromatin remodeling event that occurs in vivo during the DNA strand invasion step of HR. This ATP-dependent remodeling activity of Rad54 appears to control subsequent steps in the HR process.

Double strand breaks (DSBs) are a common form of DNA damage, resulting from a variety of environmental insults including ionizing radiation and chemical attack. The most common cause of DSBs is internal reactive oxygen species, accounting for thousands of breaks per cell per day. The inability to repair these breaks leads to genomic instability. Misrepair of these breaks can result in deletions, insertions, and translocations. In higher eukaryotes, improper DSB repair can lead to tumorigenesis. It is therefore of vital importance that cells repair these breaks accurately and faithfully.

Several pathways have evolved for the repair of DSBs. In yeast, the predominant pathway is thought to be homologous recombination (1, 2). Homologous recombination (HR) provides an "error-free" method for repairing DSBs as it utilizes homologous DNA sequences as a template to repair the lesion with no loss of genetic information. The genes in the RAD52 epistasis group, required for HR, are highly conserved from yeast to man (2–4), highlighting the importance of this molecular pathway.

Studies in yeast have suggested a sequence of molecular events that occur following the formation of a DSB (2, 4, 5). Nucleolytic end processing leads to resection of the 5' ends of DNA that flank the break, generating long stretches of 3' single-stranded DNA. Rad51p binds the single-stranded DNA, forming right-handed helical nucleoprotein filaments. In vitro, Rad52p (6), Rad54p (7), and a Rad55p/Rad57p heterodimer (8) mediate this early step by overcoming the inhibitory effects of the heterotrimeric single-stranded DNA-binding protein, RPA. In vivo, these same Rad proteins are also required for optimal recruitment of Rad51p to DNA surrounding a single DSB in yeast (7, 9). The Rad51-nucleoprotein filament is believed to search the genome for homologous sequences, resulting in the formation of a heteroduplex "joint molecule" (10). Joint molecule formation is followed by extension of the incoming strand by DNA polymerases and branch migration. The end result is the repair of the DSB without loss of genetic information (2).

Rad54p is a member of the Swi2/Snf2 family of DNA-stimulated ATPases (11), and like Swi2/Snf2, Rad54p shows ATP-dependent chromatin remodeling activity in vitro (12–14). Rad54p also binds to both single-stranded DNA (7) and Rad51p (15–17), and these latter two activities are believed to play key roles in the stimulation of Rad51-single-stranded DNA nucleoprotein filament formation. In vitro, this mediator activity of Rad54 is optimal at substoichiometric ratios of Rad54p to Rad51p (7), although in other types of in vitro assays, Rad54p functions best at stoichiometric levels (18–20). These in vitro studies raise the question of whether Rad54p might be a stoichiometric component of the extended Rad51 nucleoprotein filament in vivo.

In vitro, the ATPase activity of Rad54 is required to promote homologous DNA pairing between a Rad51-nucleoprotein filament and a double-stranded DNA or chromatin donor (12, 21). In addition, yeast harboring mutant alleles of RAD54 that disrupt the ATPase domain exhibit sensitivity to DNA-damaging agents (15) and defects in intrachromosomal recombination (21). In contrast, the ATPase-defective RAD54 alleles have little effect on interchromosomal recombination. This suggests that Rad54p may have several functions in vivo and that some, but not all, require a functioning ATPase domain. In this study, we use chromatin immunoprecipitation (ChIP) to assess the composition of the presynaptic nucleoprotein filament in vivo and to investigate the role of the Rad54 ATPase activity. We find that the ATPase activity of Rad54 is dispensable for early steps in HR but, in contrast, the ATPase activity of Rad54 is required for a chromatin remodeling step that occurs during strand invasion.

MATERIALS AND METHODS

Plasmids and Strains—The 2-μm plasmid containing the RAD54 gene plus 400 bp of upstream genomic DNA was a kind gift of Hannah Klein (New York University). A 3.2-kb NotI-SalI fragment containing the RAD54 genomic sequences was excised and cloned into the NotI.
Sall sites in pRS406. The codon for lysine 341 was mutated to arginine with QuikChange mutagenesis (Stratagene). The site-specific mutation was verified by sequencing. The mutant and wild-type plasmids were linearized with NotI and transformed into rad54Δ strains that either contained (CY1166) or lacked (CY196) silent mating type loci. All other strains used here were described previously (7).

Chromatin Immunoprecipitation—Polyclonal Rad51 and Rad52 antibodies were obtained from Santa Cruz Biotechnology. For ChIP against Rad54p and Rad55p, the proteins were tagged at their carboxyl termini with 13 copies of the e-Myc epitope as described previously (7) and immunoprecipitated with monoclonal anti-Myc (9E10) antibody (Santa Cruz Biotechnology).

ChIP assays were performed as described previously (7) with modification as follows. After adsorption to protein A-Sepharose, cross-linked antibodies were eluted in 250 μl of 2× proteinase K buffer (2× proteinase K buffer is 20 mM Tris-Cl, pH 8.0, 10 mM EDTA, 1% SDS) by shaking at 65 °C for 10 min. Beads were pelleted by pulse centrifugation, and the supernatants were diluted with an equal volume of water. Proteinase K (100 mg) was added, and samples were incubated at 42 °C for 1 h followed by 65 °C overnight. Lithium chloride (400 mM final) was added, and samples were extracted once with phenol:chloroform:iso-amyl alcohol (25:24:1) and once with chloroform:iso-amyl alcohol (24:1). Samples were then precipitated with 20 μg of glycogen and 1 ml of 100% ethanol.

Radioactive semi-quantitative PCR was performed and analyzed as described previously (7). Multiplex PCR reactions containing only the breaks-proximal (18 bp) and break-distal (900 bp) primers (see Fig. 1a) failed to amplify the expected 882-bp product even when the extension times were increased to 2 min (not shown), indicating that the DNA from the two MAT regions immunoprecipitate independently.

Micrococcal Nuclease and Southern Blotting—For micrococcal nuclease (MNase) assays, 50-ml cultures were grown in YEPl + raffinose to mid-log phase and induced with galactose as described above for ChIP assays. Parallel uninduced cultures served as negative controls. Cultures were cross-linked with formaldehyde, and extracts were prepared as for ChIP. Extracts were not sonicated but were instead treated with various amounts of MNase for 15 min at 37 °C. For optimal MNase digestion, magnesium chloride (20 μl 10 mM Tris) and calcium chloride (2 mM final) were added to the extracts. Each 50-ml culture represented one MNase digestion reaction. These reactions were quenched with 1% SDS and 5 mM EDTA and then treated with 500 μg of proteinase K at 42° for 2 h and then at 65° for 5 h to reverse the cross-links. DNA was extracted twice with phenol:chloroform:iso-amyl alcohol (25:24:1) and once with chloroform:iso-amyl alcohol (24:1) and then precipitated with sodium acetate and ethanol. Dried DNA was dissolved in 400 μl of 10 mM Tris, pH 7.4/1 mM EDTA with 300 μg RNase-A, incubated at 37° for 1 h, extracted once with phenol:chloroform:iso-amyl alcohol and then precipitated again with sodium acetate and ethanol. DNA was dissolved in water and digested with HindIII (New England Biolabs) in the appropriate buffer overnight at 37 °C. After precipitation, DNA was dissolved in 20 μl of water and electrophoresed on a 1% agarose gel. 1 μl of HindIII-DNA digest was assayed as described previously (22). The probe was made by PCR with genomic DNA as a template and primers ('5′ to 3′): GCAAGCT-TATCTCATATTAG and CTTATGTCGTTTGTTTGGTG. The location of this probe is illustrated below (see Fig. 5a). The PCR product was gel-purified (Qiagen) and labeled by the random primer method (Roche Applied Science).

For analysis of HO cleavage at HML, 25-ml cultures were grown in YEPl + raffinose and induced as described above. Formaldehyde fixation was omitted. Cells were harvested by centrifugation, washed with water, transferred to microcentrifuge tubes, and suspended in 200 μl of genomic DNA buffer (2% Triton X-100, 1% SDS, 100 mM sodium chloride, 10 mM Tris-Cl, pH = 8.0, 1 mM EDTA) and 200 μl of phenol:chloroform:iso-amyl alcohol (25:24:1) with 250 μl of zirconium beads (Biospec Products). Cells were lysed by shaking on a multivortexer for 10 min after which 200 μl of TE was added. Samples were centrifuged for 5 min, and the aqueous phase was extracted with chloroform:iso-amyl alcohol. DNA was treated with 30 μg of RNase-A for 5 min at 37° and then precipitated with sodium acetate and ethanol. Dried DNA was dissolved in water and digested overnight either with HindIII or BamHI (NEB). The DNA was precipitated, and Southern blotting was performed as described above.

RESULTS

Previously, we used chromatin immunoprecipitation analyses to monitor the recruitment of Rad proteins to a single, HO-induced DSB at the yeast MAT locus (7). Although our studies focused on recruitment events immediately proximal to the DSB, Haber and colleagues (9) demonstrated that Rad51p is cross-linked over several kilobases of DNA surrounding the DSB in yeast. To determine whether Rad54p and the other members of the RAD52 group are part of this extended nucleoprotein filament, chromatin immunoprecipitation analyses were performed for Rad51p, Rad52p, Rad54p, and Rad55p using DNA primer sets that monitor recruitment events both proximal to (18–257 bp) and distal from the induced DSB (Fig. 1a). We also included primers to the PHO5 locus as a control in the multiplex PCR analysis. Consistent with previous studies (7, 9), we observe rapid recruitment of Rad51p to DNA both proximal to and distal from the induced DSB (Fig. 1, b and c). Likewise, we find that Rad52p and Rad55p are also recruited to both regions, consistent with their association with the entire Rad51p nucleoprotein filament (Fig. 1, b and c). In contrast, Rad54p is only cross-linked to DNA proximal to the DSB (Fig. 1, b and c). Identical results are found in ChIP time courses that are extended to 8 h (data not shown). Thus, these data indicate that Rad54p is not a stoichiometric component of a Rad51 nucleoprotein filament in vivo, and furthermore, the data indicate that Rad54p may only associate with the 3′ end of the presynaptic complex.

Previously, we showed that Rad52p, Rad55p, and Rad54p are all mediators of Rad51p recruitment to DNA proximal to an HO-induced DSB in vivo (7). Given that Rad54p is only associated with this same proximal region, we tested whether the mediator activity of Rad54 is restricted to the 3′ end of the presynaptic filament. As we showed previously, deletion of RAD54 causes a 6-fold decrease in recruitment of Rad51p to DNA proximal to the HO-induced DSB (Fig. 2, a and b). However, cross-linking of Rad51p at the distal region (600–900 bp) is reduced less than 2-fold in the rad54 deletion strain. Likewise, binding of Rad51p to a medial region (250–535 bp, Fig. 2b) is reduced only 3-fold in the absence of Rad54p. In contrast, deletion of the RAD52 gene leads to a complete absence of Rad51p recruitment throughout the entire region tested (Fig. 2b). Thus, these data distinguish the mediator activities of Rad54p and Rad52p, and they indicate that the Rad54p mediator activity is most crucial in the same region where Rad54p itself binds, at the extreme 3′ end of the presynaptic filament.

Based on analogy to prokaryotic homologous recombination (4), the Rad51p presynaptic filament is believed to search for homologous sequences until it is able to form an unstable paracentric joint in which homologous sequences are aligned, but the DNA strands are not topologically intertwined. This paranemic joint in which homologous sequences are aligned, until it is able to form an unstable paranemic joint in which homologous sequences are aligned, but the DNA strands are not topologically intertwined.

This unstable joint is subsequently extended to the 3′ end of the Rad51p filament, forming a stable plectonemic DNA joint. Previously, we showed that RAD54 is required for cross-linking of Rad51p to homologous donor sequences, and at that time, we concluded that Rad54p might play a role in the homology search process (7). However, this analysis only monitored binding of Rad51p to donor sequences that are homologous to the extreme 3′ end of the incoming presynaptic filament (Fig. 3; HML proximal), a region that we have now shown is devoid of Rad51p in a rad54 deletion strain. Thus, it is possible that an unstable paracentric joint might be formed in the absence of RAD54, given the high levels of Rad51p that are bound to the distal end of the presynaptic filament.

Fig. 3b shows a representative Rad51p ChIP analysis in strains harboring a galactose-inducible HO gene that also contain the homologous donor sequences, HML, and HMR. PCR analysis of Rad51p immunoprecipitates included primer sets for the PHO5 control locus, the MAT locus, and the HML homologous donor locus. For HML, two sets of primer pairs...
were used: 1) a proximal \textit{HML} set that detects synapsis of the 3' end of the incoming filament and 2) a distal \textit{HML} set that detects synapsis of the medial region of the presynaptic filament (Figs. 2b and 3a). As we observed previously (7), Rad51p is detected at the \textit{MAT} locus (Fig. 3b) and at the proximal \textit{HML} donor site in the wild-type strain (not shown). However, detection of Rad51p in the distal region of \textit{HML} was not observed even in the wild-type strain unless we dramatically decreased the amount of sonication during the ChIP procedure (Fig. 3, b and c). Importantly, decreasing the extent of sonication had little impact on the immunoprecipitation efficiency of \textit{MAT} sequences in the wild-type strain. In the \textit{rad54} deletion strain, Rad51p was not detectable at the proximal \textit{HML} region (not shown), confirming our previous results (7). However, Rad51p...
was seen bound to the downstream HML region in the rad54 deletion strain when sonication cycles were decreased (Fig. 3, b and c). Thus, these data indicate that RAD54 is not required for a successful homology search, but rather, that the absence of Rad54p leads to the formation of an unstable DNA joint that may only be parancemic in nature. A similar conclusion has been made by Sugawara et al. (9).

Rad54 is unique among those proteins known to mediate the binding of Rad51p to single-stranded DNA as Rad54 is a potent DNA-stimulated ATPase. Several roles have been proposed for the ATPase activity of Rad54, including ATP-dependent DNA duplex separation during strand invasion (19, 21, 23), ATP-dependent removal of Rad51p at a post-synthesis step (24), as well as ATP-dependent chromatin remodeling during the homology search or DNA strand invasion (12, 25). To identify ATP-dependent role(s) for Rad54p during HR, we created isogenic yeast strains that express wild-type Rad54p or an ATPase-defective version of Rad54, rad54K341R. Similar to previous studies (1, 15), both the rad54 deletion strain and the strain that expresses the ATPase-defective version, rad54K341R, are hypersensitive to the DNA-damaging agent, methylmethanesulfonate (data not shown).

ChIP analyses were performed to monitor Rad51p recruitment in the isogenic RAD54, rad54K341R, and rad54 deletion strains. In the isogenic strain set that lacks homologous donors, Rad51p was detected at both the MAT proximal and the MAT distal regions in the RAD54 strain (Fig. 4a). When the rad54K341R allele was expressed, Rad51p recruitment to the proximal region was restored. Thus, this role for Rad54p in facilitating Rad51p recruitment does not require an intact ATPase domain. Furthermore, in a similar isogenic strain set that harbors homologous donors, both Rad54p and Rad54K341R supported binding of Rad51p to the HML proximal region, even under normal sonication conditions (Fig. 4b). Thus, the ATPase-defective version of Rad54p is able to rescue full synapsis at the HML donor. However, only strains harboring wild-type Rad54p were able to extend the DNA joint and generate a strand invasion/extension (SIE) product (Fig. 4c).

These data demonstrate that Rad54p has at least two functions during recombinational DNA repair: 1) a presynaptic activity that helps recruit Rad51p to single-stranded DNA and is independent of ATP hydrolysis and 2) a subsequent activity, dependent on an intact ATPase domain, which is required for extension of the incoming strand by DNA polymerases.
ase activity in the enhanced accessibility of HML donor chromatin, we treated yeast with hydroxyurea for several hours to deplete nucleotide pools, block DNA polymerase activity, and arrest cells in early S phase. Galactose was then added to the arrested cells to induce the formation of the DSB at MAT. As shown in Fig. 5c, hydroxyurea treatment has no effect on the efficiency of DSB formation at the MAT locus, but hydroxyurea did abolish the formation of a 30-bp strand invasion/extension product at HML. Notably, hydroxyurea treatment had only a minor effect (−30% decrease) on the HO cleavage at HML (Fig. 5d). Thus, the increased accessibility of HML chromatin does not require extension of the DNA strand by DNA polymerases, but the enhanced accessibility to HO appears to reflect a RAD54-dependent step that occurs after completion of a successful homology search. These data are consistent with a model in which the nucleosome that is positioned over the HO cleavage site inhibits the formation of the completely intertwined DNA joint, and ATP-dependent chromatin remodeling by Rad54p overcomes this barrier, facilitating the formation of a stable DNA joint that can be extended by DNA polymerases.

**DISCUSSION**

We have previously demonstrated that Rad54p has chromatin remodeling activity *in vitro* (12). Here, we present the first...
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In this case, Rad54p requires an intact ATPase domain to remodel at least one nucleosome positioned over the HO cleavage site at the HML donor locus. In vitro, the ATP-dependent remodeling activity of Rad54p leads to enhanced accessibility of nucleosomal sites to restriction enzymes, but it does not appear to lead to dramatic changes in the positioning of nucleosomes within array substrates (12). Likewise, in vivo, we were unable to detect significant changes in nucleosome positioning by MNase analysis during the early stages of strand invasion, even when chromatin states were fixed with formaldehyde. Thus, the data are consistent with the idea that ATP-dependent remodeling by Rad54 may lead to enhanced accessibility of nucleosomal DNA without large scale nucleosome movements.

Previous studies have suggested that Rad54p has both ATP-dependent and -independent functions in vivo (1, 21, 24, 28). Here, we have confirmed these observations and identified the functions. The presynaptic activity of Rad54p does not require an intact ATPase domain, and consequently, a sonication-sensitive DNA joint is formed at a donor locus in the presence of an ATPase-defective version of Rad54p. The ATP-dependent activity of Rad54p appears to comprise, at least in part, an ATP-dependent chromatin remodeling function that facilitates the formation of a stable DNA joint that can be acted on by DNA polymerases. Whether there exist additional ATP-dependent steps for Rad54p remains to be investigated. The dual functionality of Rad54p may have evolved as genomes became larger and more highly condensed chromatin structures began to interfere with HR. A mutation that disrupts the ATPase domain of human RAD54 is found in some tumors (29), suggesting that ATP-dependent chromatin remodeling by Rad54 may play a key role in maintaining genome integrity among all eukaryotes.

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REFERENCES

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